

POULTRY LITTER ASH AS A PHOSPHORUS SOURCE FOR GREENHOUSE CROP  
PRODUCTION

A Dissertation

Submitted to the Graduate Faculty of  
Louisiana State University and  
Agricultural and Mechanical College  
in partial fulfillment of the  
requirements for the degree of  
Doctor of Philosophy

in

The School of Plant, Environmental and Soil Sciences

by  
Daniel Wells  
B.S., Auburn University 2006  
M.S., Auburn University 2008  
May 2013

# Effects of Phosphorus Source, Phosphorus Rate, and Liming Rate on Growth and Quality of *Verbena canadensis* Britton ‘Homestead Purple’ and *Lantana camara* L. ‘New Gold’<sup>1</sup>

Daniel E. Wells<sup>2</sup>, Edward W. Bush<sup>3</sup>, Jeffrey S. Beasley<sup>3</sup>, and Charles E. Johnson<sup>4</sup>

School of Plant, Environmental, and Soil Sciences, LSU AgCenter

Louisiana State University and A & M College

104 Sturgis Hall, Baton Rouge, LA 70803

## Abstract

Phosphate rock ores, rich in phosphorus (P), are mined in great quantities around the world for the production of P fertilizers. However, availability of phosphate reserves is in question due to decreasing global supplies. Therefore, application of alternative, renewable P sources is of interest. Poultry litter ash (PLA), a byproduct of bioenergy production, contains P concentrations comparable to conventional fertilizers. In this experiment, two P sources, PLA and superphosphate (SP), were compared at two P application rates and two liming rates as fertilizer sources for the production of two commonly-grown greenhouse crops (*Verbena canadensis* Britton ‘Homestead Purple’ and *Lantana camara* L. ‘New Gold’). Application of PLA produced plants of comparable biomass and quality to those fertilized with SP. Increasing P application rate, across both P sources, increased total flower numbers 42 and 26% for verbena and lantana, respectively. Foliar P concentrations of verbena and lantana increased 27 and 62% for PLA-fertilized plants compared to SP. In addition, substrate pH increased 25% using PLA versus SP. Overall, PLA supplies adequate P fertility and does not reduce pH compared to the more water-soluble, rock phosphate based SP fertilizer.

**Index words:** phosphorus, poultry litter ash, *Verbena canadensis* Britton ‘Homestead Purple’, *Lantana camara* L. ‘New Gold’, dolomitic limestone.

**Species used in this study:** *Verbena canadensis* Britton ‘Homestead Purple’; *Lantana camara* L. ‘New Gold’.

## Significance to the Nursery Industry

Due to declining global and domestic phosphate rock ore reserves, application of alternative, renewable phosphorus (P) fertilizers is of great importance. Poultry litter ash (PLA) performed as well as or better than superphosphate (SP) in terms of growth, biomass, quality, and P-fertilization of greenhouse-grown, containerized verbena and lantana. Substrate pH was increased 25% when PLA was used instead of SP, while substrate EC was comparable. Foliar P concentrations were higher for plants fertilized with PLA than those fertilized with SP. Foliar Mn concentrations were greatly reduced through PLA application compared with SP, while foliar Ca concentrations were slightly increased. Results indicate that PLA is a suitable alternative to SP, a phosphate rock ore-based fertilizer. In addition, PLA incorporation increases substrate pH, potentially reducing dependency on commonly used liming amendments. Poultry litter ash may reduce costs associated with P fertilization and substrate liming, while also reducing dependency on mined phosphate rock ore-based fertilizers. Future research should address P dissolution rates from PLA in soilless substrates.

## Introduction

Phosphorus (P) is the eleventh most abundant element in the earth’s crust and is essential for most life forms, including plants (36). Phosphorus fertilizers commonly used in

plant production systems, are mined and processed from phosphate-rich ore deposits in the earth’s crust (25). However, future availability of phosphate rock ore reserves is in jeopardy (12). According to Roberts and Stewart (30), the United States’ phosphate rock ore reserves, at current production, are estimated at less than 20 years. Current global commercial phosphate reserves are estimated to be depleted within 50 to 100 years (12). While expected durations of global and domestic phosphate reserves and resources are only estimates, a decline in phosphate rock ore quality is a consensus among speculators and scientists (12, 30, 36, 39). Therefore, development of renewable, high quality P fertilizer sources is of paramount importance.

Historically, animal manures have been utilized as fertilizers in production of many agricultural commodities. Given concerns regarding phosphate rock ore for P fertilizers, manures have gained interest as potential recycled P sources (13). The rise in poultry production has resulted in significant amounts of waste being produced from these facilities in many areas of the United States, particularly in the southeastern U.S. Poultry litter is a biomass source consisting predominantly of bird manure and bedding materials (31). Bedding materials typically consist of straw, sawdust, wood shavings, shredded paper, peanut hulls, and/or rice hulls, depending on location and availability of materials (22). Poultry litter contains comparable amounts of nitrogen to ruminant wastes, but higher concentrations of P, since fowls are unable to extract organically bound-P from feeds with the addition of phytase (38). Like most manures, poultry litter application as a fertilizer source is limited due to high transportation costs (7) and environmental concerns associated with surface water impairment (35).

To alleviate environmental concerns due to geographical-concentrated poultry litter applications as well as expand

<sup>1</sup>Received for publication January 28, 2013; in revised form February 14, 2013.

<sup>2</sup>Graduate student. dwel111@lsu.edu.

<sup>3</sup>Associate professor.

<sup>4</sup>Professor.

the use of poultry litter as a recyclable fertilizer source, several methods have been employed to reduce weight of, or concentrate P within, poultry litter including, compaction (7), pelletization (24), composting (9), P removal (40), gasification (27), and combustion (10, 34). Of all these methods, combustion of poultry litter may be the most efficient means available because the ash contains inorganic P while energy released during combustion could be used for electricity or heat production (19). Although the combustion process can be more complicated for poultry litter compared to traditional fuel sources due to inconsistent composition, moisture content, and high ash content (6), combustion of poultry litter is technologically feasible (16). For example, Fibrominn power plant in Benson, MN, an alternative energy plant, co-combusts poultry litter and wood to provide energy to approximately 40,000 homes. Ash from the combustion process is sold as a commercial fertilizer (26) with the majority of ash a product of combusted poultry litter (19, 21).

With the increase in capability of using poultry litter for ashing or power production the potential of poultry litter ash (PLA) as a fertilizer source needs to be examined for a variety of cropping systems. Limited scientific experiments have reported PLA is a suitable nutrient source for several agronomic crops including wheat (*Triticum aestivum* L.) (10), Japanese mustard spinach (*Brassica rapa* L.) (14), buckwheat (*Fagopyrum esculentum* L.) (14), oil radish (*Raphanus sativus oleiformis* Adagio), phacelia (*Phacelia tanacetifolia* Lisette), or ryegrass (*Lolium multiflorum westerwoldicum* Gordo) (5). In each of these experiments, researchers reported increased plant P accumulation for soils amended with PLA even though PLA-P is characterized as having low water solubility (11, 5). No experiments have examined PLA as a P source for containerized horticultural crops.

Given the uncertainty of future P ore based fertilizer availability and quality, low cost alternatives such as PLA may be highly desirable in nursery and greenhouse production systems that require high P fertilization additions with high water usage. Additionally, environmental concerns due to P losses from highly concentrated production sites may be reduced by utilizing less soluble, recycled P sources. Therefore, the objective of the experiment was to examine the use of PLA as an alternative P source during the production of two commonly-grown greenhouse crops (*Lantana camara* L. 'New Gold' and *Verbena canadensis* Britton 'Homestead Purple').

## Materials and Methods

**Experiment design.** Eighty *Lantana camara* L. 'New Gold' and *Verbena canadensis* Britton 'Homestead Purple' plants growing in 105-cell trays were selected for uniform quality and size for the experiment initiated February 3, 2012. For each species, two plants were transplanted into 1.6-liter containers for a total of 40 containers per species. Containers were filled with a substrate composed of pine bark (<0.38 cm) and peat moss (4:1 by vol) and pre-plant incorporated with 0.89 kg·m<sup>-3</sup> of micronutrient package (Micromax, Scotts Company, Marysville, OH), and 0.25 kg K·m<sup>-3</sup> (0N-0P-35.7K). Remaining pre-plant amendments were superphosphate (SP; 18% P<sub>2</sub>O<sub>5</sub>) or poultry litter ash (PLA; 7% P<sub>2</sub>O<sub>5</sub>), incorporated at 140 or 280 g P·m<sup>-3</sup>, in combination with pulverized dolomitic limestone (DL) at 1.5 or 3.0 g·m<sup>-3</sup>. Poultry litter ash used in this experiment was the product of commercial energy production via combustion of

poultry litter and was obtained courtesy of North American Fertilizer, LLC. Containers filled with the eight pre-plant incorporated combinations were arranged in a completely randomized design with five single-container replications. All plants were maintained under greenhouse conditions at an average temperature of 27.7C (82F), with no supplemental irradiance, for 42 and 70 days for *Verbena canadensis* and *Lantana camara*, respectively. During the experiment, plants were supplied with 350 ml water per day including 120 ml aliquots per container N at 250 mg NH<sub>4</sub>NO<sub>3</sub>·liter<sup>-1</sup>·d<sup>-1</sup>.

**Plant response.** Plant growth was measured bi-weekly using a growth index [(height + widest width + perpendicular width) / 3] and flower number was quantified for flower buds showing color. Leaf samples, composed of the most recently matured leaves, were removed, dried at 60C (140F) for 72 hours, and biomass recorded before tissue was milled to <0.5 mm using a Thomas Wiley® Mini-Mill (Thomas Scientific, Swedesboro, NJ). Tissue was digested in concentrated nitric acid at an average of 120C (250F), diluted to 20 ml with deionized water, and filtered prior to analysis of elemental Al, B, Ca, Cu, Fe, Mg, Mn, Mo, P, K, Na, S, and Zn concentrations using inductively coupled plasma optical emission spectroscopy (SPECTRO Analytical Instruments, Kleve, Germany; Louisiana State University Soil Testing and Plant Analysis Laboratory, Baton Rouge, LA). At 42 and 70 days, plant shoots were harvested at the substrate surface, dried at 60C (140F) for 72 hours, and biomass recorded.

**Leachate collection and analysis.** Leachate samples measuring 90 ml, from three containers per treatment for *Lantana camara*, were collected bi-weekly following the Virginia Tech extraction method (41). Leachate samples were transported to the laboratory and allowed to cool to room temperature [21C (70F)] prior to leachate-pH and EC measurement (Orion Star A215 solution analyzer; Thermo Scientific Inc., Beverly, MA).

**Statistical analysis.** The experiment was a completely randomized design with five replications. Growth index, flower counts, plant dry weight, leachate pH, EC, and tissue nutrient analyses data were analyzed following the mixed procedure in SAS/STAT® statistical software (32). Means for each measurement at each collection interval were separated using Tukey's Honest Significant Difference Test at a significance level of 0.05.

## Results and Discussion

**Plant response of verbena.** *Verbena* growth was not significantly influenced by P source, P rate, or DL rate at 14 or 28 DAP (Table 1). At 42 DAP *verbena* growth, measured using a growth index, increased 9.5% from 40.0 to 44.2 across both P sources as DL rate increased from 1.5 to 3.0 kg·m<sup>-3</sup>. However, in the case of shoot dry weight, DL affected *verbena* growth differently depending on P source and rate of application (Table 2). Increasing DL rate from 1.5 to 3.0 kg·m<sup>-3</sup> did not increase *verbena* shoot dry weights at 21.2 g and 23.2 g, respectively, in combination with the lower SP application rate. However, increasing DL application rate at the higher SP application rate of 280 g P·m<sup>-3</sup> resulted in higher dry weight of 30.4 g compared to 19.8 g. For PLA, increasing the DL application rate increased *verbena* shoot dry weight 5.9 g at the lower PLA application rate. In fact,

**Table 1.** Effects of phosphorus rate and dolomitic limestone rate on growth index of *Verbena canadensis* ‘Homestead Purple’ and *Lantana camara* ‘New Gold’ over experimental periods of 42 and 70 days, respectively.

P source <sup>z</sup>	P rate (g·m <sup>-3</sup> )	DL rate <sup>y</sup> (kg·m <sup>-3</sup> )	Verbena			Lantana				
			Growth index <sup>x</sup>							
			14 DAP <sup>w</sup>	28 DAP	42 DAP	14 DAP	28 DAP	42 DAP	56 DAP	70 DAP
—	140	—	25.5a <sup>v</sup>	35.9a	42.1a	15.3a	22.9a	36.4a	45.8a	51.3a
—	280	—	26.7a	33.3a	42.1a	16.3a	25.6a	33.1a	42.5b	47.3a
—	—	—	NS <sup>u</sup>	NS	NS	NS	NS	NS	0.029	NS
—	—	1.5	25.1a	33.6a	40.0b	15.3a	24.1a	31.8b	41.1b	46.8b
—	—	3.0	27.1a	35.7a	44.2a	16.3a	24.5a	37.7a	47.2a	51.8a
			NS	NS	0.048	NS	NS	0.013	0.001	0.047

<sup>z</sup>SP = superphosphate; PLA = poultry litter ash.

<sup>y</sup>DL = pulverized dolomitic limestone.

<sup>x</sup>Growth index was measured in cm as: [(Height + Widest Width + Perpendicular Width) / 3].

<sup>w</sup>Days after potting.

<sup>v</sup>Values in column followed by different letters are significant according to Tukey’s Studentized Range Test ( $\alpha = 0.05$ ).

<sup>u</sup>P-value derived from analysis of variance; NS = not significant.

the combination of DL at 3.0 kg·m<sup>-3</sup> and PLA at 140 g·m<sup>-3</sup> resulted in shoot dry weight of 26.6 g comparable to 27.3 and 28.1 g as the DL rate increased at 280 g P·m<sup>-3</sup> of PLA. An increasing effect of DL on plant dry weight was not evident at the higher PLA application rate.

Phosphorus application rate, regardless of P source, had the greatest effect on verbena flower counts throughout the experiment (Table 3). Flower counts increased from 7.4 to 12, 12.8 to 19.2, and 11.1 to 13.3, at 14, 28, and 42 DAP, respectively, as the rate of P increased from 140 to 280 g·m<sup>-3</sup>. Over the 42-day experiment, total flower counts increased 42% from 31.3 to 44.5 as P rate increased from 140 to 280 g·m<sup>-3</sup>. In general, verbenas fertilized with 280 g P·m<sup>-3</sup> across all DL rates resulted in greater flowering than verbena fertilized at 140 g P·m<sup>-3</sup>.

**Table 2.** Effects of phosphorus source, phosphorus rate, and dolomitic lime rate on shoot dry weights of *Verbena canadensis* ‘Homestead Purple’ and *Lantana camara* ‘New Gold’ harvested at 42 and 70 days after potting, respectively.

P source <sup>z</sup>	P rate (g·m <sup>-3</sup> )	DL rate <sup>y</sup> (kg·m <sup>-3</sup> )	Shoot dry weight (g)	
			Verbena	Lantana
SP	—	1.5	20.5c <sup>x</sup>	24.9b
SP	—	3.0	26.8a	28.9a
PLA	—	1.5	23.9b	29.2a
PLA	—	3.0	27.3a	26.6ab
			0.0217 <sup>v</sup>	0.0029
SP	140	1.5	21.2c	25.9ab
SP	280	1.5	19.8c	24.0b
SP	140	3.0	23.2bc	27.9ab
SP	280	3.0	30.4a	29.8ab
PLA	140	1.5	20.6c	26.9ab
PLA	280	1.5	27.3a	31.6a
PLA	140	3.0	26.5ab	25.4ab
PLA	280	3.0	28.1a	27.9ab
			<0.0001	NS

<sup>z</sup>SP = superphosphate; PLA = poultry litter ash.

<sup>y</sup>DL = pulverized dolomitic limestone.

<sup>x</sup>Values in column followed by different letters are significant according to Tukey’s Studentized Range Test ( $\alpha = 0.05$ ).

<sup>v</sup>P-value derived from analysis of variance; NS = not significant.

*Plant response of lantana.* As noted with verbena, P source did not affect lantana growth. However, lantana growth index was affected by DL rate and P rate (Table 1). Lantana growth index increased from 31.8 to 37.7, 41.1 to 47.2, and 46.8 to 51.8, at 42, 56, and 70 DAP, respectively, as DL rate increased from 1.5 to 3.0 kg·m<sup>-3</sup>. In general, P application rate did not affect growth index of lantana with the exception of a 7% decrease at 56 DAP when P rate was increased. Shoot dry weight of lantana was not singularly affected by P source, P rate, or DL rate (Table 2). However, increasing DL rate from 1.5 to 3.0 kg·m<sup>-3</sup> increased shoot biomass from 24.9 to 28.9 g of lantanas fertilized with SP, but did not affect those fertilized with PLA.

Similar to verbena, flower counts of lantana were affected by P application rate throughout the experiment (Table 3). Flower counts increased from 48.8 to 66.8, 110.8 to 150.3, 106.8 to 116.3, and 93.8 to 123.7, at 28, 42, 56, and 70 DAP, respectively, when P application rate of either P source was increased from 140 to 280 g P·m<sup>-3</sup>. For the experiment, there was an overall increase of 26%, from 382.8 to 483.5 flowers, when P application rate was increased. Similar to shoot dry weight, increasing DL rate from 1.5 to 3.0 kg·m<sup>-3</sup>, increased total flower counts of lantana fertilized with SP from 387.7 to 469.8, but did not affect flower counts of lantanas fertilized with PLA.

*Substrate leachate-pH and EC from lantana camara.* Substrate leachate-pH was affected at every measurement date by P source and DL rate (Table 4). Average leachate-pH increased 25%, from pH 5.18 to 6.48, when the P source was changed from SP to PLA, and 9.3%, from pH 5.57 to 6.09, when DL rate was increased 1.5 to 3.0 kg DL·m<sup>-3</sup>. As DL rate increased from 1.5 to 3.0 kg DL·m<sup>-3</sup>, substrate leachate-pH increased an average of 11%, from 4.87 to 5.48, for plants fertilized with SP, but only 6%, from 6.27 to 6.69, for those fertilized with PLA.

Substrate leachate-EC was also affected by P source at 0, 7, 14, 21, 49, and 63 DAP (Table 5). Leachate-EC was highest when plants were fertilized with PLA at 0, 7, 49, and 63 DAP, but was higher for SP-fertilized plants 14 and 21 DAP. Increasing the P application rate from 140 g P·m<sup>-3</sup>

**Table 3. Effects of phosphorus source, phosphorus rate, and dolomitic limestone rate on bi-weekly and cumulative flower counts of *Verbena canadensis* ‘Homestead Purple’ and *Lantana camara* ‘New Gold’ over experimental periods of 42 and 70 days, respectively.**

P source <sup>y</sup>	P rate (g·m <sup>-3</sup> )	DL rate <sup>x</sup> (kg·m <sup>-3</sup> )	Flower count <sup>z</sup>									
			Verbena				Lantana					
			14 DAP <sup>w</sup>	28 DAP	42 DAP	Total	14 DAP	28 DAP	42 DAP	56 DAP	70 DAP	Total
—	140	—	7.4b <sup>y</sup>	12.8b	11.1b	31.3b	22.8a	48.8b	110.8b	106.8b	93.8b	382.8b
—	280	—	12.0a	19.2a	13.3a	44.5a	26.5a	66.8a	150.3a	116.3a	123.7a	483.5a
			<0.0001 <sup>u</sup>	<0.0001	0.0492	<0.001	NS	<0.0001	<0.0001	0.0391	0.0012	<0.0001
SP	—	1.5	9.0a	13.5a	10.7a	33.2b	18.8b	52.2b	113.8b	102.0a	100.8a	387.7b
SP	—	3.0	11.0a	15.7a	12.8a	39.5ab	32.0a	65.5a	147.0a	114.0a	111.3a	469.8a
PLA	—	1.5	10.2a	17.3a	12.7a	40.2a	23.8b	57.2ab	118.8b	114.8a	114.3a	429.0ab
PLA	—	3.0	8.7a	17.3a	12.7a	38.7ab	23.8b	56.2ab	142.3a	115.3a	108.3a	446.0a
			0.0311	NS	NS	0.0276	0.0276	0.0259	NS	NS	NS	0.0105
SP	140	1.5	7.0c	11.0d	10.3a	28.3b	17.7b	47.3cd	96.3d	98.3a	94.7a	354.3c
SP	280	1.5	11.0abc	16.0abcd	11.0a	38.0ab	20.0b	57.0bcd	131.3c	105.7a	107.0a	421.0bc
SP	140	3.0	7.7bc	12.3cd	13.0a	33.0b	29.7ab	53.7bcd	128.0c	114.0a	97.3a	422.7bc
SP	280	3.0	14.3a	19.0abc	12.7a	46.0a	34.3a	77.3a	166.0a	114.0a	125.3a	517.0a
PLA	140	1.5	8.0bc	13.3bcd	11.7a	33.0b	23.7ab	49.3bcd	94.0d	106.7a	98.0a	371.7c
PLA	280	1.5	12.3ab	21.3a	13.7a	47.3a	24.0ab	65.0abc	143.7bc	123.0a	130.7a	486.3ab
PLA	140	3.0	7.0c	14.3abcd	9.3a	30.7b	20.0b	44.7d	124.7c	108a	85.0a	382.3c
PLA	280	3.0	10.3abc	20.3ab	16.0a	46.7a	27.7ab	67.7ab	160.0ab	122.7a	131.7a	509.7a
			NS	NS	NS	NS	NS	NS	NS	NS	NS	NS

<sup>z</sup>Flower buds showing color at the time of data collection.

<sup>y</sup>SP = superphosphate; PLA = poultry litter ash.

<sup>x</sup>DL = pulverized dolomitic limestone.

<sup>w</sup>Days after potting.

<sup>y</sup>Values in column followed by different letters are significant according to Tukey’s Studentized Range Test ( $\alpha = 0.05$ ).

<sup>u</sup>P-value derived from analysis of variance; NS = not significant.

**Table 4. Effects of phosphorus source and dolomitic lime rate on substrate leachate-pH measured weekly from *Lantana camara* ‘New Gold’ over an experimental period of 63 days.**

P source <sup>z</sup>	P rate (g·m <sup>-3</sup> )	DL rate <sup>y</sup> (kg·m <sup>-3</sup> )	Substrate leachate-pH										Average
			0 DAP <sup>x</sup>	7 DAP	14 DAP	21 DAP	28 DAP	35 DAP	42 DAP	49 DAP	56 DAP	63 DAP	
SP	—	—	4.33b <sup>w</sup>	4.65b	4.79b	4.89b	5.03b	5.17b	5.49b	5.69b	5.82b	5.89b	5.18b
PLA	—	—	5.93a	6.27a	6.31a	6.52a	6.59a	6.68a	6.68a	6.57a	6.57a	6.65a	6.48a
			<0.0001 <sup>v</sup>	<0.0001	<0.0001	<0.0001	<0.0001	<0.0001	<0.0001	<0.0001	<0.0001	<0.0001	<0.0001
—	—	1.5	4.93b	5.19b	5.30b	5.44b	5.53b	5.63b	5.81b	5.86b	5.90b	6.06b	5.57b
—	—	3.0	5.32a	5.72a	5.80a	5.97a	6.10a	6.22a	6.36a	6.39a	6.49a	6.48a	6.09a
			<0.0001	<0.0001	<0.0001	<0.0001	<0.0001	<0.0001	<0.0001	<0.0001	<0.0001	<0.0001	<0.0001
SP	—	1.5	4.05d	4.30d	4.44d	4.62d	4.74d	4.87d	5.16d	5.36d	5.48d	5.65d	4.87d
SP	—	3.0	4.61c	4.99c	5.14c	5.17c	5.33c	5.46c	5.82c	6.03c	6.16c	6.13c	5.48c
PLA	—	1.5	5.81b	6.08b	6.16b	6.27b	6.33b	6.39b	6.46b	6.37b	6.32b	6.47b	6.27b
PLA	—	3.0	6.05a	6.45a	6.45a	6.77a	6.87a	6.97a	6.90a	6.77a	6.82a	6.84a	6.69a
			0.0004	<0.0001	<0.0001	NS	NS	NS	0.028	0.0001	0.0035	NS	NS
SP	140	1.5	4.17e	4.41f	4.49f	4.70d	4.74c	4.94f	5.22f	5.58d	5.62d	5.84f	4.97ef
SP	280	1.5	3.94e	4.19f	4.38f	4.52d	4.74e	4.80f	5.10f	5.13e	5.33e	5.46g	4.76f
SP	140	3.0	4.78c	5.16d	5.44d	5.57c	5.54c	5.69d	6.00d	6.22b	6.25bc	6.07ef	5.67d
SP	280	3.0	4.43d	4.83e	4.85e	4.78d	5.11d	5.23e	5.65e	5.84c	6.07c	6.18de	5.30e
PLA	140	1.5	5.78b	6.02c	6.11c	6.16b	6.27b	6.21c	6.33c	6.38b	6.23bc	6.39cd	6.19c
PLA	280	1.5	5.83ab	6.13bc	6.22bc	6.37ab	6.38b	6.56b	6.60bc	6.36b	6.41b	6.55bc	6.34bc
PLA	140	3.0	6.06a	6.33ab	6.34b	6.77a	6.84a	6.89a	6.85ab	6.71a	6.88a	6.78ab	6.64ab
PLA	280	3.0	6.04a	6.58a	6.57a	6.76a	6.91a	7.05a	6.94a	6.83a	6.76a	6.89a	6.73a
			NS	NS	0.0002	NS	0.0152	NS	NS	NS	0.0012	0.0011	NS

<sup>z</sup>SP = superphosphate; PLA = poultry litter ash.

<sup>y</sup>DL = pulverized dolomitic limestone.

<sup>x</sup>Days after potting.

<sup>w</sup>Values in column followed by different letters are significant according to Tukey’s Studentized Range Test ( $\alpha = 0.05$ ).

<sup>v</sup>P-value derived from analysis of variance; NS = not significant.

**Table 5. Effects of phosphorus source and phosphorus rate on substrate leachate-EC measured from *Lantana camara* ‘New Gold’.**

P source <sup>z</sup>	P rate (g·m <sup>-3</sup> )	Substrate leachate-EC										
		0 DAP <sup>y</sup>	7 DAP	14 DAP	21 DAP	28 DAP	35 DAP	42 DAP	49 DAP	56 DAP	63 DAP	Average
SP	—	2.24b <sup>x</sup>	2.51b	2.48a	1.98a	1.38a	1.10a	0.85a	0.66b	0.66a	0.57b	1.44a
PLA	—	2.92a	2.99a	2.36b	1.86b	1.39a	1.09a	0.82a	0.73a	0.70a	0.66a	1.55a
		<0.0001 <sup>w</sup>	<0.0001	0.0017	0.04	NS	NS	NS	0.0043	NS	0.0112	NS
—	140	2.36b	2.34b	2.10b	1.60b	1.13b	0.91b	0.74b	0.59b	0.58b	0.56b	1.29b
—	280	2.80a	3.16a	2.75a	2.25a	1.64a	1.28a	0.93a	0.80a	0.78a	0.67a	1.71a
		<0.0001	<0.0001	<0.0001	<0.0001	<0.0001	<0.0001	<0.0001	<0.0001	<0.0001	0.0031	0.0015
SP	140	2.04d	2.12d	2.14c	1.77b	1.29c	0.99c	0.79bc	0.59c	0.59c	0.52b	1.28b
SP	280	2.44c	2.91b	2.84a	2.20a	1.49b	1.20b	0.90ab	0.74b	0.78a	0.62ab	1.60ab
PLA	140	2.68b	2.56c	2.06c	1.43c	0.97d	0.82d	0.68c	0.59c	0.57c	0.60ab	1.30b
PLA	280	3.16a	3.42a	2.66b	2.30a	1.80a	1.36a	0.96a	0.86a	0.82a	0.71a	1.81a
		NS	NS	NS	0.001	<0.0001	0.0001	0.0312	0.0036	0.0027	NS	NS

<sup>z</sup>SP = superphosphate; PLA = poultry litter ash.

<sup>y</sup>Days after potting.

<sup>x</sup>Values in column followed by different letters are significant according to Tukey’s Studentized Range Test ( $\alpha = 0.05$ ).

<sup>w</sup>P-value derived from analysis of variance; NS = not significant.

to 280 g P·m<sup>-3</sup> than for at every measurement date and was increased by an average of 33% for the 70 day experiment. While leachate-EC was affected by both P source and P application rate, increasing P rate from 140 to 280 g P·m<sup>-3</sup> increased leachate-EC for plants fertilized with PLA by a higher margin than for those fertilized with SP at 21, 28, 35, 42, 49, and 56 DAP.

*Foliar nutrient concentrations in verbena.* Foliar concentrations of Ca, Mn, and P were affected by P source for

verbenas (Table 6). Foliar Ca and P concentrations increased from 0.61 to 0.78% and 0.26 to 0.33%, respectively, when verbenas were fertilized with PLA compared to SP, while Mn concentrations decreased from 122.97 to 65.01 mg·kg<sup>-1</sup>. A similar trend was also exhibited, across both P sources, as foliar P increased from 0.27 to 0.31%, and foliar Mn decreased from 106.01 to 81.96 mg·kg<sup>-1</sup> with P application rate increase from 140 to 280 g P·m<sup>-3</sup>. Across both P sources and P rates, increasing DL rate from 1.5 to 3.0 kg·m<sup>-3</sup> decreased foliar Mn concentrations from 100.21 to 87.77 mg·kg<sup>-1</sup>. At

**Table 6. Effects of phosphorus source, phosphorus rate, and dolomitic lime rate on foliar nutrient concentrations of *Verbena canadensis* ‘Home-stead Purple’ and *Lantana camara* ‘New Gold’.**

P source <sup>z</sup>	P rate (g·m <sup>-3</sup> )	DL rate <sup>y</sup> (kg·m <sup>-3</sup> )	Verbena					Lantana				
			Ca <sup>x</sup>	Mg	Mn	P	K	Ca	Mg	Mn	P	K
SP	—	—	0.61b <sup>w</sup>	0.52a	122.97a	0.26b	1.89a	0.56b	0.32b	257.50a	0.21b	1.24a
PLA	—	—	0.78a	0.49a	65.01b	0.33a	1.90a	0.80a	0.38a	130.02b	0.34a	1.46a
			0.0072 <sup>v</sup>	NS	<0.0001	<0.0001	NS	<0.0001	<0.0001	<0.0001	<0.0001	NS
—	140	—	0.68a	0.52a	106.01a	0.27b	1.88a	0.65b	0.31b	204.67a	0.24b	1.43a
—	280	—	0.71a	0.48a	81.96b	0.31a	1.92a	0.71a	0.40a	182.84b	0.31a	1.27a
			NS	NS	<0.0001	0.0014	NS	0.0021	<0.0001	0.0024	0.0003	NS
—	—	1.5	0.69a	0.49a	100.21a	0.29a	1.88a	0.64b	0.33b	184.54b	0.28a	1.38a
—	—	3.0	0.70a	0.51a	87.77b	0.29a	1.92a	0.73a	0.37a	202.98a	0.27a	1.32a
			NS	NS	0.0063	NS	NS	0.0001	<0.0001	0.0077	NS	NS
SP	140	1.5	0.51a	0.50a	131.66a	0.26bc	1.98ab	0.45e	0.22e	241.36b	0.18c	1.56a
SP	280	1.5	0.60a	0.48a	126.53a	0.27bc	2.02ab	0.53de	0.36bcd	255.88ab	0.25bc	1.22a
SP	140	3.0	0.71a	0.61a	145.53a	0.22c	1.71ab	0.66bc	0.32d	291.06a	0.16c	1.09a
SP	280	3.0	0.64a	0.47a	88.16b	0.31ab	1.86ab	0.60cd	0.38bc	241.70b	0.26bc	1.07a
PLA	140	1.5	0.82a	0.52a	87.61b	0.32ab	1.84ab	0.71bc	0.33d	138.70cd	0.29b	1.32a
PLA	280	1.5	0.84a	0.46a	55.04c	0.33ab	1.67b	0.85a	0.40b	102.20d	0.39a	1.41a
PLA	140	3.0	0.69a	0.45a	59.26c	0.30ab	1.99ab	0.77ab	0.35cd	147.58c	0.33ab	1.75a
PLA	280	3.0	0.77a	0.51a	58.13c	0.35a	2.11a	0.88a	0.45a	131.58cd	0.33ab	1.37a
			NS	NS	<0.0001	NS	NS	NS	0.0006	0.0031	0.0416	NS

<sup>z</sup>SP = superphosphate; PLA = poultry litter ash.

<sup>y</sup>DL = pulverized dolomitic limestone.

<sup>x</sup>Macronutrients reported as percentage of dry matter. Mn reported in mg·kg<sup>-1</sup> dry matter.

<sup>w</sup>Values in columns followed by different letters were significant according to Tukey’s Honest Significance Difference Test ( $\alpha = 0.05$ ).

<sup>v</sup>P-value derived from analysis of variance; NS = not significant.

the high DL rate, increasing P rate from 140 to 280 g·m<sup>-3</sup> decreased foliar Mn concentration from 145.53 to 88.16 mg·kg<sup>-1</sup> for verbenas fertilized with SP, but did not affect foliar Mn concentrations of verbenas fertilized with PLA.

*Foliar nutrient concentrations in lantana.* For lantanas, foliar Ca, Mg, Mn, and P concentrations were affected by P source and P application rate (Table 6). When PLA was used as the P source, foliar Ca, Mg, and P increased from 0.56 to 0.80%, 0.32 to 0.38%, and 0.21 to 0.34%, respectively. However, similar to verbenas, foliar Mn concentrations were decreased from 257.5 to 130.02 mg·kg<sup>-1</sup> when PLA was the P source. The same general trend existed for P application rate. As P rate increased from 140 to 280 g·m<sup>-3</sup> foliar Ca, Mg, and P concentrations increased from 0.65 to 0.71%, 0.31 to 0.40%, and 0.24 to 0.31%, respectively, while foliar Mn concentrations decreased from 204.67 to 182.84 mg·kg<sup>-1</sup>. When P source was SP and the DL rate was highest, foliar Mn concentrations decreased from 291.06 to 241.70 mg·kg<sup>-1</sup> as P rate increased from 140 to 280 g·m<sup>-3</sup>, but Mn concentrations were not affected when PLA was applied.

Poultry litter ash is an acceptable P source for verbenas and lantana greenhouse container production compared to water-soluble, phosphate rock ore-based fertilizers such as SP. Verbenas and lantana growth, measured using a growth index, and in terms of biomass, exhibited similar patterns to plants fertilized using SP. Codling (10) reported similar results for wheat (*Triticum aestivum* L.) grown on two differing soil types when comparing PLA to potassium phosphate as P fertilizer sources. Similarly, in an experiment conducted to determine the effects of PLA on soil-P pools and P uptake, Bachmann and Eichler-Lobermann (5) reported no differences in biomass per species of buckwheat (*Fagopyrum esculentum* L.), oil radish (*Raphanus sativus oleiformis* Adagio), phacelia (*Phacelia tanacetifolia* Lisette), or ryegrass (*Lolium multiflorum westerwoldicum* Gordo) when comparing PLA and potassium phosphate. In addition, flower count, a common measurement used for ornamental plant quality, increased as P rate increased for PLA and SP. James and Van Iersel (20) reported flower numbers, of other ornamental species, to be positively affected by increasing P fertility. Bi et al. (8) reported increased flower numbers for marigold with increasing poultry litter application rates. Therefore, under the conditions tested for ornamental plant production, PLA, a low water soluble P source, was able to provide adequate P concentrations throughout the 42 and 70 day production cycles to result in marketable quality plants without any observable deleterious effects.

Although PLA was primarily examined for its suitability as a P source, PLA also affected substrate pH known to influence nutrient availability and uptake. Research has shown substrate amendments such as fertilizers and pH-adjusting materials can greatly affect plant growth and quality as a direct result of changes in substrate chemical properties (3, 4, 37). Unlike SP, which is known to reduce substrate pH (18) and require higher lime additions to maintain a range of optimal pH, PLA did not lower substrate pH. In fact, verbenas and lantana growth, within species, was similar for PLA across both the lower and higher DL rates.

Poultry litter ash contains a high concentration of Ca as a result of litter composition prior to ashing that can result in a high alkaline compound (11). Although substrate leachate-pH often exceeded the recommended range of 5.4

to 6.8 for proper plant growth (15) during the experiment, P plant uptake was not negatively affected at the higher DL in combination with PLA. Solution pH-dependent dissociation constants for H<sub>3</sub>PO<sub>4</sub> of 2.1 and 7.2 suggest the monovalent P species (H<sub>2</sub>PO<sub>4</sub><sup>-</sup>) available for plant uptake would not be affected within the pH ranges measured for PLA-fertilized plants during the course of the experiment (33). Therefore, PLA has the added benefit of adjusting media pH while supplying P that should reduce liming requirements of soilless substrates.

Because PLA is not a pure P source and contains constituents that can affect substrate chemical properties and crop nutrition (11, 5), effects of PLA incorporation on salt leaching and ancillary nutrient uptake should be characterized. Substrate EC was not affected by DL rate, but was affected by P source and P rate, with P rate having the most consistent effect. Exceedingly high substrate EC was not observed with PLA incorporation. Leachate-EC measurements generally remained within an optimal range of 0.5 to 3.0 mS·cm<sup>-1</sup> (28, 29) throughout the experiment, with the only exceptions occurring at the highest rate of PLA at 0 and 7 DAP. In general, PLA did not affect the availability or uptake of required nutrients other than P. However, Ca uptake was increased and Mn uptake was decreased for plants fertilized with PLA. Increased Ca uptake was most likely due to increased Ca concentrations as a result of PLA application. More interesting were the changes in foliar Mn across P sources, P rates, and DL rates. In the case of SP-fertilized plants, Mn uptake increased due to higher Mn availability at lower substrate pH compared to plants fertilized with the more alkaline PLA. Although plant Mn toxicities occur in organic soils and soilless substrates, toxicity levels have been shown to be ameliorated with applications of Fe (17), K (1), Ca (2), or Mg (23). Manganese toxicity symptoms were not observed in this experiment likely due to Fe, Ca, K, and Mg being supplied in adequate concentrations.

Substrate pH was affected throughout the experiment and substrate EC was increased for the first week due to PLA incorporation, but adequate concentrations of P were supplied to plants verbenas and lantana. However, based on data recorded for this experiment, it is unknown what salts contributed to leachate-EC. High concentrations of P have been shown to be rapidly released from SP in soilless substrates (42), but P dissolution rates from PLA in a soilless substrate have not previously been reported. Continued research should determine the rate and concentration of P dissolution from PLA when used as a P source in a soilless substrate.

## Literature Cited

1. Alam, S., F. Akiha, S. Kamei, S. Imamul Huq, and S. Kawai. 2005. Mechanism of potassium alleviation of manganese phytotoxicity in barley. *J. Plant Nut.* 28:889–901.
2. Alam, S., R. Kodama, F. Akiha, S. Kamei, and S. Kawai. 2006. Alleviation of manganese phytotoxicity in barley with calcium. *J. Plant Nutr.* 29:59–74.
3. Altland, J.E. and M.G. Buamscha. 2008. Nutrient availability from Douglas fir bark in response to substrate pH. *HortScience* 43:478–483.
4. Argo, W.R. and J.A. Biernbaum. 1996. The effect of lime, irrigation-water source, and water-soluble fertilizer on root-zone pH, electrical conductivity, and macronutrient management of container root media with impatiens. *J. Amer. Soc. Hort. Sci.* 121:442–452.
5. Bachmann, S. and B. Eichler-Lobermann. 2010. Soil phosphorus pools as affected by application of poultry litter ash in combination with catch crop cultivation. *Commun. Soil Sci. Plan.* 41:1098–1111.

6. Baranyai, V. and S. Bradley. 2008. Turning Chesapeake bay watershed poultry manure and litter into energy: An analysis of the impediments and the feasibility of implementing energy technologies in the Chesapeake bay watershed in order to improve water quality. Chesapeake Bay Program: A Watershed Partnership. CBP/TRS-289-08.
7. Bernhart, M., O.O. Fasina, C.W. Wood, and J. Fulton. 2010. Compaction of poultry litter. *Biores. Tech.* 101:234–238.
8. Bi, G., W.B. Evans, J.M. Spiers, and A.L. Witcher. 2010. Effects of organic and inorganic fertilizers on marigold growth and flowering. *HortScience* 45:1373–1377.
9. Brodie, H.L., L.E. Carr, and P. Condon. 2000. A comparison of static pile and turned windrow methods for poultry litter compost production. *Compost Sci. Util.* 8:178–189.
10. Codling, E.E., R.L. Chaney, and J. Sherwell. 2002. Poultry litter ash as a potential phosphorus source for agricultural crops. *J. Environ. Qual.* 31:954–961.
11. Codling, E.E. 2006. Laboratory characterization of extractable phosphorus in poultry litter and poultry litter ash. *Soil Science*. 171:858–864.
12. Cordell, D., J. Drangert, and S. White. 2009. The story of phosphorus: Global food security and food for thought. *Global Environmental Change* 19:292–305.
13. Dawson, C.J. and J. Hilton. 2011. Fertiliser availability in a resource-limited world: Production and recycling of nitrogen and phosphorus. *Food Policy* 36:S14–S22.
14. Faridullah, M.I., S. Yamamoto, A.E. Eneji, T. Uchiyama, and T. Honna. 2009. Recycling of chicken and duck litter ash as a nutrient source for Japanese mustard spinach. *J. Plant Nutr.* 32:1082–1091.
15. Fonteno, W.C., P.V. Nelson, and D.A. Bailey. 1996. Plug substrates: In search of the perfect mix. The systems approach to growing plugs: Water, substrate, and nutrition. NCSU Plug Research and Information Development Center, N.C. State Univ.
16. Habetz, D. and R. Echols. 2006. Development of successful poultry litter-to-energy furnace. ASABE Paper No. 064185. St. Joseph, MI. ASABE.
17. Handreck, K.A. 1997. Low iron supply aggravates manganese toxicity in zonal geraniums growing in soilless potting media. *J. Plant Nutr.* 20:1593–1605.
18. Huang, J. and P.V. Nelson. 2001. Impact of pre-plant root substrate amendments on soilless substrate EC, pH, and nutrient availability. *Commun. Soil Sci. Plan* 32:2863–2875.
19. IPEP. 2006. North Alabama Integrated Poultry and Ethanol Production (IPEP) Feasibility Study: Final Report for NRCS Grant Agreement 68-3A75-3-144. Prepared by T.R. Miles Technical Consultants, Inc. and B.R. Bock Consulting, Inc.
20. James, E. and M. Van Iersel. 2001. Ebb and flow production of petunias and begonias as affected by fertilizers with differing phosphorus content. *HortScience* 36:282–285.
21. Jia, L. and E.J. Anthony. 2011. Combustion of poultry-derived fuel in a coal-fired pilot-scale circulating fluidized bed combustor. *Fuel Process. Technol.* 92:2138–2144.
22. Kelleher, B.P., J.J. Leahy, A.M. Henihan, T.F. O'Dwyer, D. Sutton, and M.J. Leahy. 2002. Advances in poultry litter disposal technology — a review. *Biores. Tech.* 83:27–36.
23. Le Bot, J., M.J. Goss, G.P.R. Carvalho, M.L. Van Beusichem, and E.A. Kirkby. 1990. The significance of the magnesium to manganese ratio in plant tissues for growth and alleviation of manganese toxicity in tomato (*Lycopersicon esculentum*) and wheat (*Triticum aestivum*) plants. *Plant and Soil* 124:205–210.
24. McMullen, J., O.O. Fasina, C.W. Wood, and Y. Feng. 2005. Storage and handling characteristics of pellets from poultry litter. *Applied Eng. Agric.* 21:645–651.
25. Mikkelsen, R.L. and T.W. Bruulsema. 2005. Fertilizer use for horticultural crops in the U.S. during the 20<sup>th</sup> century. *HortTechnology* 15:24–30.
26. Misra, M.K., K.W. Ragland, and A.J. Baker. 1993. Wood ash composition as a function of furnace temperature. *Biomass and Bioenergy* 4:103–116.
27. Priyadarsan, S., S. Mukhtar, M.T. Holtzapple, K. Annamalai, and J.M. Sweeten. 2004. Fixed-bed gasification of feedlot manure and poultry litter biomass. *Trans. of the ASAE.* 47:1689–1696.
28. Raviv, M. and H. Lieth. 2008. *Soilless Culture: Theory and Practice.* Elsevier, Oxford, UK.
29. Robbins, J.A. and M.R. Evans. 2005. Growing media for container production in a greenhouse or nursery. Part II (Physical and Chemical Properties). Univ. Ark. Coop. Ext. Serv. FSA 6098-PD-4-04R.
30. Roberts, T.L. and W.M. Stewart. 2002. Inorganic phosphorus and potassium production and reserves. *Better Crops* 86(2):6–7.
31. Robinson, J.S. and A.N. Sharpley. 1996. Reaction in soil of phosphorus released from poultry litter. *Soil Sci. Soc. Amer. J.* 60:1583–1588.
32. SAS Institute Inc. 2011. SAS/STAT® 9.3 User's Guide. SAS Institute, Inc. Cary, NC.
33. Schachtman, D.P., R.J. Reid, and S.M. Ayling. 1998. Phosphorus uptake by plants: From soil to cell. *Plant Physiol.* 116:447–453.
34. Schiemenz, K. and B. Eichler-Lobermann. 2010. Biomass ashes and their phosphorus fertilizing effect on different crops. *Nutr. Cycl. Agroecosyst.* 87:471–482.
35. Sharpley, A.N., S.C. Chapra, R. Wedepohl, J.T. Sims, T.C. Daniel, and K.R. Reddy. 1994. Managing agricultural phosphorus for protection of surface waters: Issues and options. *J. Environ. Qual.* 23:437–451.
36. Smil, V. 2000. Phosphorus in the environment: Natural flows and human interferences. *Annu. Rev. Energy Environ.* 25:53–88.
37. Smith, B.R., P.R. Fisher, and W.R. Argo. 2004. Water-soluble fertilizer concentration and pH of a peat-based substrate affect growth, nutrient uptake, and chlorosis of container-grown seed geraniums. *J. Plant Nutr.* 27:497–524.
38. Sommers, L.E. and A.L. Sutton. 1980. Use of waste materials as sources of phosphorus. *In: The Role of Phosphorus in Agriculture.* F.E. Khasawneh et al., eds. Am. Soc. Agron., Madison, WI.
39. Steen, I. 1998. Phosphorus availability in the 21<sup>st</sup> century: Management of a non-renewable resource. *Phosphorus and Potassium* 217:25–31.
40. Szogi, A.A., M.B. Vanotti, and P.G. Hunt. 2008. Phosphorus recovery from poultry litter. *Trans. ASABE* 51:1727–1734.
41. Wright, R.D. 1986. The pour-thru nutrient extraction procedure. *HortScience* 21:227–229.
42. Yeager, T.H. and J.E. Barrett. 1984. Phosphorus leaching from <sup>32</sup>P-superphosphate-amended soilless container media. *HortScience* 19:216–217.